Diploneis (Ehrenberg) Cleve 1894

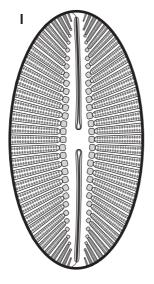
Type species: Diploneis didyma (Ehrenberg) Cleve

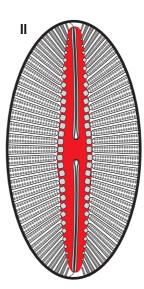
Characteristics – Cells biraphid, elliptical with bluntly rounded apices, sometimes constricted mid-valve (Fig. 133: C). Longitudinal canals (II) parallel to the raphe, striae composed of complex (loculate) areolae, usually clearly visible under LM (Fig. 133: E-G; Fig. 134: D). When observed under SEM the longitudinal canals are perforated on the exterior of the valve with areolae but not on the interior (Fig. 134: C). Cells heavily silicified.

Plastid structure – Cells with two plastids, one on either side of the apical plane, may be many lobed (Fig. 133: A, C, D) or simple (Fig. 133: B).

Identification of species – Species can be identified by cell size, cell shape and structure and density of the striae as well as the shape of the axial and central area and the presence and degree of the mid-valve constriction. Some species have unique structures such as square openings in the longitudinal canals (Fig. 134:A).

Ecology – Cells solitary and motile. Freshwater forms found in the benthos of acidic oligotrophic waters as well as alkaline waters with higher trophic status and conductivity. Also found in moist sub-aerial habitats.





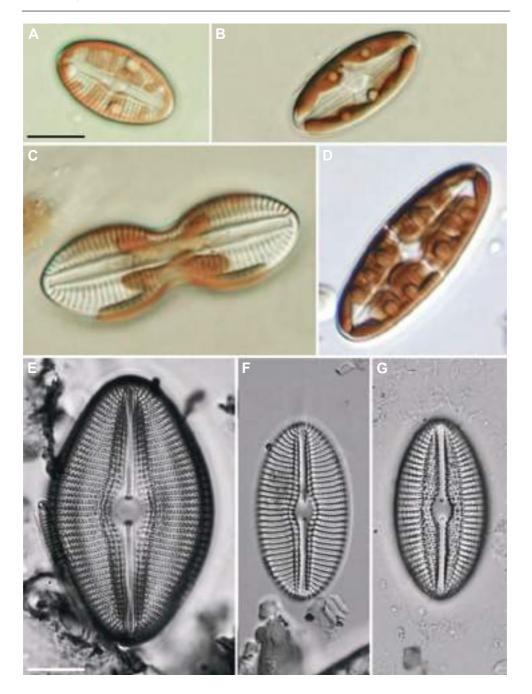


Fig. 133. Diploneis spp. A-G. LM. A-D. Living cells, valve views showing a variety of plastid shapes. E-F. Valve views of cleaned material.
G. Valve view of Diploneis fenestrata J.C. Taylor & B. Karthick. Scale bars = 10 μm (A-G).

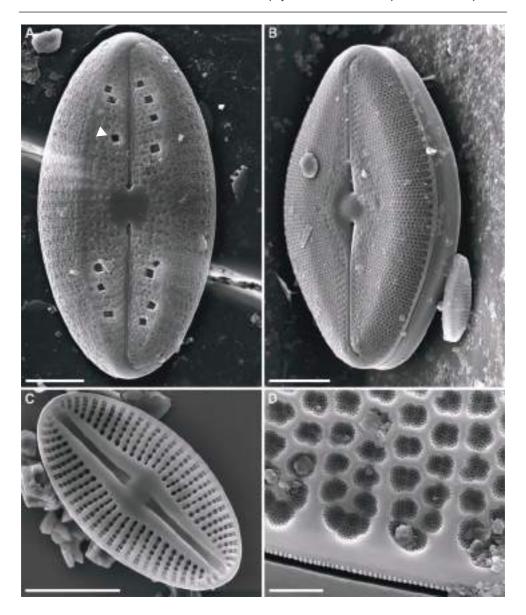


Fig. 134. *Diploneis* spp. **A-D.** SEM. **A.** *Diploneis fenestrata*, valve view, note square openings in longitudinal canals (arrow). **B.** *Diploneis* sp., oblique view of valve exterior. **C.** *Diploneis* sp., view of valve interior. **D.** Exterior view, detail of complex loculate areolae.

Scale bars = $5 \mu m$ (A), 10 μm (B-C), 1 μm (D).

Adlafia Gerd Moser, Lange-Bertalot & Metzeltin 1998

Type species: *Adlafia muscora* (Kociolek & Reviers) Gerd Moser, Lange-Bertalot & Metzeltin

SYNONYM:

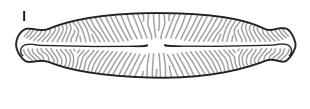
Navicula Bory 1822 pro parte

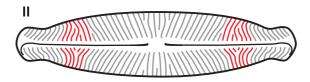
Characteristics – Cells biraphid, small, striae can often be rather fine and difficult to resolve in LM. Striae mid-valve are strongly radiate and curved becoming strongly convergent towards the apices (II). Cells may have slightly undulating margins (Fig. 135: C) and in some cases the areolae may be visible (Fig. 135: C-D). The closely related *Kobayasiella* (see Fig. 141) usually has denser striation and a small bend or kink about halfway along the length of the raphe which is not found in *Adlafia*.

Plastid structure – Single plastid with two lobes connected by a bridge (H-shape) (Fig. 135: B).

Identification of species – Species in this genus are distinguished based on cell size and shape and the shape of the apices. Striae density and angle relative to the **transapical axis** are also important characteristics to consider.

Ecology – Cells solitary. Found in acidic oligotrophic waters and moist sub-aerial habitats.





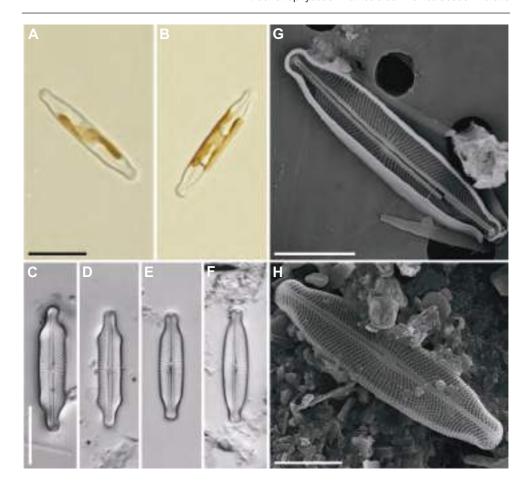


Fig. 135. *Adlafia* spp. **A-F.** LM. **A-B.** Living cells, valve views. **C-F.** Valve views of cleaned material. **G-H.** SEM. **G.** Internal view of valve. **H.** External view of valve. Scale bars = $10 \mu m$ (A-F), $5 \mu m$ (G-H).

Capartogramma Kufferath 1956

Type species: Capartogramma jeanii Kufferath

SYNONYM:

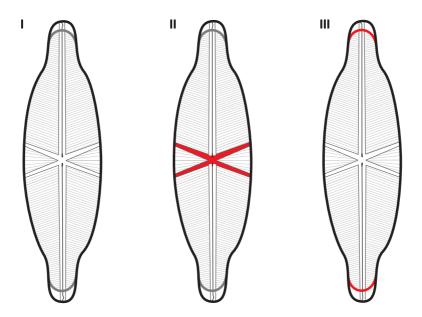
Schizostauron Grunow 1867 pro parte Stauroneis Ehrenberg 1843 pro parte

Characteristics – Cells **biraphid** with fine striae, areolae small and difficult to observe under LM. This genus is characterised by X-shaped silica thickening in the central area (II, Fig. 136: L) and a **pseudoseptum** (III) which is present at each apex. Axial area very narrow. Cells usually bilaterally symmetrical, however *Capartogramma amphoroides* R. Ross has a dorsiventral symmetry.

Plastid structure – Cells with one large plastid (Fig. 136: A).

Identification of species – Species in this genus are distinguished based on cell size, cell symmetry and shape, and the shape of the apices.

Ecology – Cells solitary and motile. Found in the benthos and plankton, with greatest species diversity in tropical African waters



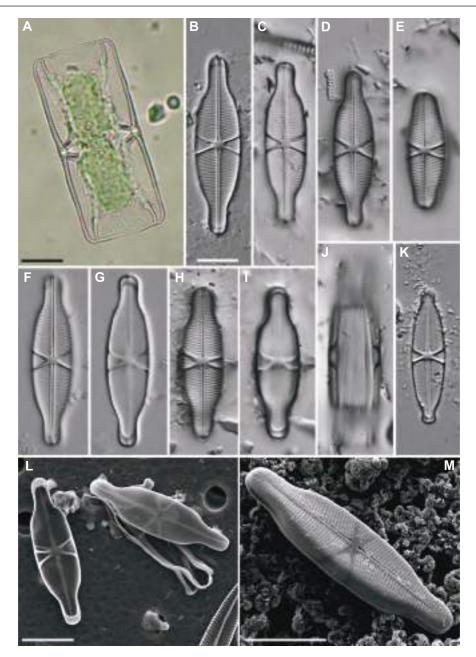


Fig. 136. Capartogramma spp. A-K. LM. A. Living cell of *C. karstenii* (O. Müller) R. Ross, girdle view. B-J. *C. crucicula* (Grunow ex R. Cleve) Ross; valve views at various foci (B-I.); girdle view (J). K. Valve view of *C. crucicula* [var. parva Fusey]. L-M. SEM. L. External view of valve and copulae (right); internal view of valve (left). M. External view of valve showing valve mantle. Scale bars = 10 μm.

Eolimna Lange-Bertalot & W. Schiller 1997

Type species: Eolimna martinii W. Schiller & Lange-Bertalot

SYNONYM:

Navicula Bory 1822 pro parte

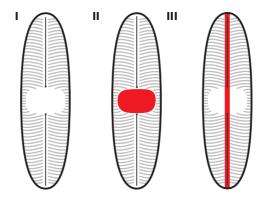
Characteristics – Cells **biraphid**, very small, elliptical to linear elliptical with broadly rounded apices. Striae fine, radiate or parallel composed of single rows of areolae which are not discernable under LM. Raphe straight and simple (I). Central area (II) variable in size but never extending to the valve margins. Axial area very narrow (III).

Plastid structure – Cells with one plastid which may be simple (Fig. 137: A) or lobed (Fig. 137: C).

Identification of species – Species can be identified by cell size, cell shape, shape of the apices, structure and density of the striae as well as shape of the central area.

Ecology – Cells solitary, free living and motile. Found in the benthos of oligotrophic to eutrophic waters in both low and moderate conductivities.

Note – We have provided a description and illustration of the genus *Eolimna* to demonstrate the recent (last decade) concept of this genus. This genus was originally described from fossil material. Recently small naviculoid diatoms ascribed to *Eolimna* have been examined in terms of plastid structure and genetic relationships and it has been concluded that the majority of the small species we currently consider *Eolimna* (e.g. *Eolimna minima* (Grunow) Lange-Bertalot) should be included with *Selllaphora* or in other genera such as *Craticula* (e.g. *Craticula subminuscula* (Manguin) Wetzel & Ector).



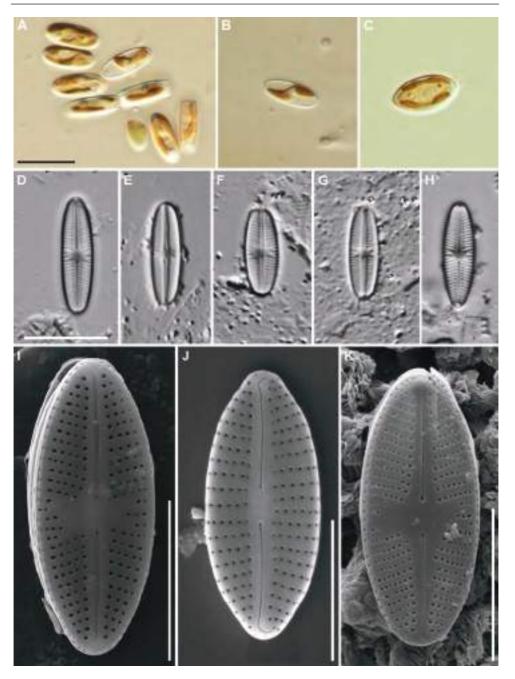


Fig. 137. Eolimna spp. A-H. LM. A-B. Living cells, valve and girdle views. C. Living cell, valve view of Eolimna subminuscula (Manguin) Gerd Moser, Lange-Bertalot & Metzeltin. D-H. Eolimna sp., valve view. I-K. SEM. I, K. External view of valve of Eolimna sp. J. E. subminuscula, external view of valve.

Scale bars = 10 μm (A-H), 5 μm (I-K).

Fistulifera Lange-Bertalot 1997

Type species: Fistulifera saprophila (Lange-Bertalot & Bonik) Lange-Bertalot

SYNONYM:

Navicula Bory 1822 pro parte

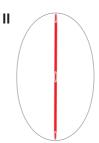
Characteristics – Cells **biraphid**, very small, elliptical with broadly rounded apices. Striae not discernable under LM (Fig. 138: A-D) and also difficult to resolve with SEM (Fig. 138: E). Raphe straight and simple (Fig. 138: A-D) carried in a sternum which is usually the only structure which can be seen using LM (II). Slight swellings present in the sternum at the central nodule and apices.

Plastid structure – Cells with one H-shaped plastid with 2 large lipid bodies.

Identification of species – Up till now only one species known from tropical Africa: *Fistulifera saprophila.*

Ecology – Cells solitary, free living and motile. Found in the benthos of eutrophic to hypereutrophic waters with moderate to high conductivities.





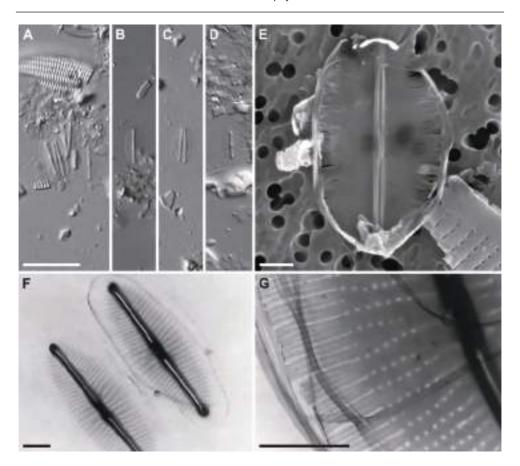


Fig. 138. Fistulifera saprophila. A-D. LM, valve views. E. SEM, internal view of valve. F-G. Transmission electron microscopy. Scale bars = 10 μ m (A-D), 1 μ m (E-G).

Geissleria Lange-Bertalot & Metzeltin 1996

Type species: Geissleria moseri Metzeltin, Witkowski & Lange-Bertalot

SYNONYM:

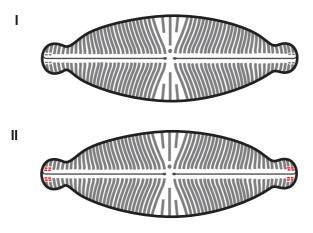
Navicula Bory 1822 pro parte

Characteristics – Cells biraphid, elliptical to linear elliptical with capitate to broadly rounded apices. Raphe straight. Striae parallel in the mid-valve becoming radiate and often curved (Fig. 139), and parallel to convergent at the apices. Areolae are discernable under LM. Isolated punctum often present in the central area (Fig. 139: D-E). Chief distinguishing characteristic of this genus is the presence of annulae at the poles (II). Annulae are 1-4 transapical striae, often composed of areolae with a distinctive structure, which interrupt the striae.

Plastid structure – Not observed in tropical African material.

Identification of species – Species can be identified by cell size, cell shape, shape of the apices, and orientation, curvature and density of the striae as well as shape of the central area. Presence/absence and position of an isolated punctum and the structure of the **annulae**.

Ecology – Cells solitary, free living and motile. Found in the benthos of oligotrophic to eutrophic waters in both low and moderate conductivities.



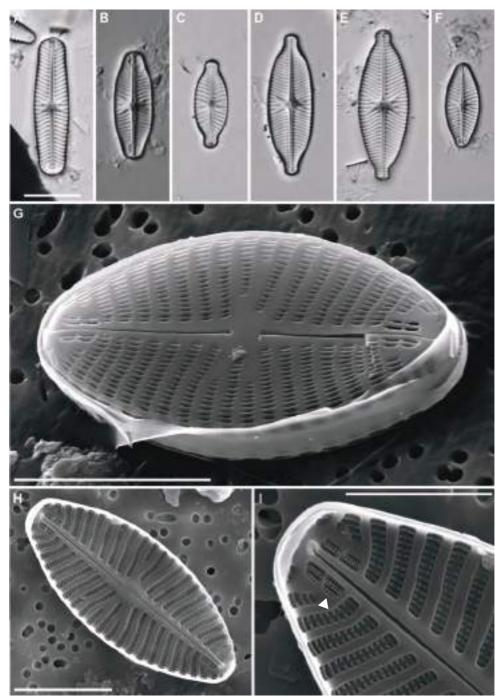


Fig. 139. *Geissleria* spp. A-F. LM, valve views of various species. G-I. SEM. G. External view of valve. H-I. Internal view of valve. I. Detail of annulae (arrow). Scale bars = 10 μ m (A-F), 5 μ m (G-H), 3 μ m (I).

Hippodonta Lange-Bertalot, Metzeltin & Witkowski 1996

Type species: *Hippodonta lueneburgensis* (Grunow) Lange-Bertalot, Metzeltin & Witkowski

SYNONYM:

Navicula Bory 1822 pro parte

Characteristics – Cells biraphid, small, elliptical to linear elliptical with broadly rounded, rounded or subcapitate apices. Striae very robust composed of double, rarely single rows of areolae which are usually not discernable under LM. Raphe straight and simple (Fig. 140: C-P), terminal endings do not extend onto the valve mantle. Thickened bars of silica present at the poles (II; Fig. 141: F) on the valve interior.

Plastid structure – Two plastids one each side of the cell next to the girdles (Fig. 140: A-B).

Identification of species – Species can be identified by cell size, cell shape, shape of the apices, orientation and density of the striae as well as structure of the central area.

Ecology – Cells solitary, free living and motile. Found in the benthos of oligotrophic to eutrophic waters in both low and moderate conductivities.





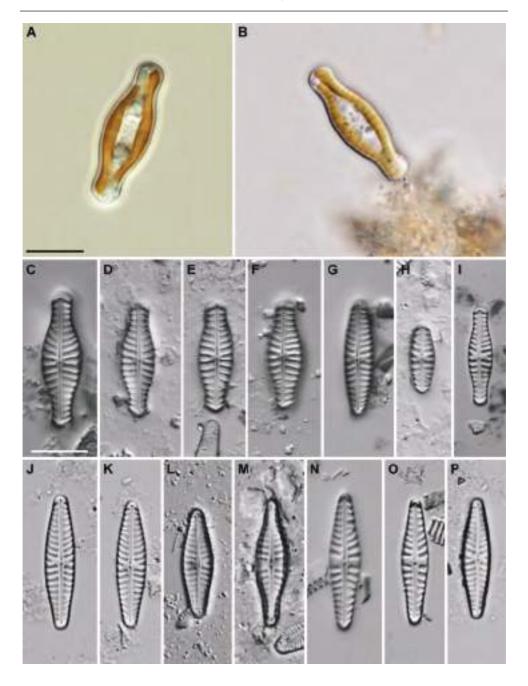


Fig. 140. Hippodonta spp. A-F. LM. A-B. Living cells of Hippodonta capitata (Ehrenberg) Lange-Bertalot, Metzeltin & Witkowski. B-P. Cleaned valves. G-H. Hippodonta hungarica (Grunow) Lange-Bertalot, Metzeltin & Witkowski. Scale bar = 10 μm (A-P).

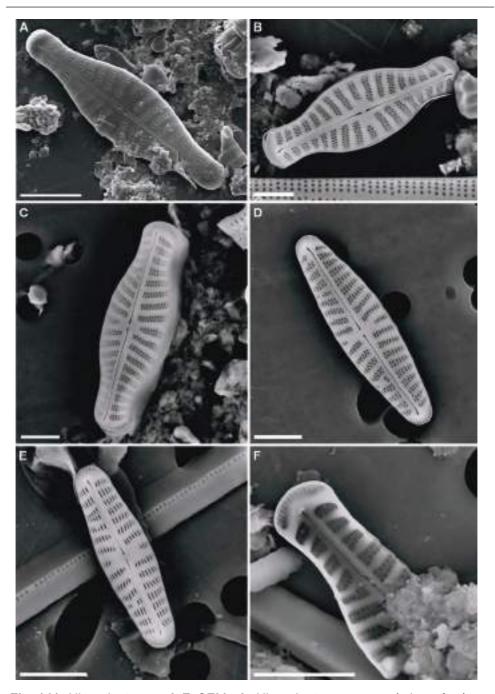


Fig. 141. *Hippodonta* spp. **A-F.** SEM. **A.** *Hippodonta* sp., external view of valve. **B-C.** *H.* capitata, external view of valve. **D-E.** *Hippodonta* spp., external view of valves. **F.** *H.* capitata, internal view of valve, note thickened bar of silica at apex. Scale bars = $5 \mu m$ (A, E-F), $4 \mu m$ (C-D), $2 \mu m$ (B).

Kobayasiella Lange-Bertalot 1999

Type species: Kobayasiella bicuneus (Lange-Bertalot) Lange-Bertalot

SYNONYM:

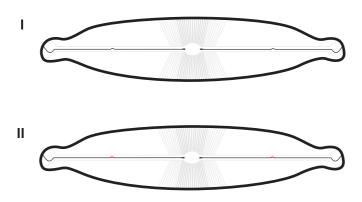
Navicula Bory 1822 pro parte *Kobayasia* Lange-Bertalot 1996

Characteristics – Cells biraphid, mostly linear elliptical in shape with broadly subcapitate, capitate or rostrate apices. Striae very fine, composed of areolae difficult to resolve even under SEM, radiate at mid-valve becoming abruptly convergent near the apices. Raphe straight and simple (Fig. 142: B-F) with a characteristic undulation or kink approximately halfway along the length of the raphe branch (II; Fig. 142: G). Central area variable in size, usually small but may be slightly expanded. Axial area very narrow.

Plastid structure – Single plastid with 2 lobes connected by one or more bridges (Fig. 142: A).

Identification of species – Species can be identified by cell size, cell shape, shape of the apices, orientation and density of the striae as well as structure of the central area.

Ecology – Cells solitary, free living and motile. Found in the benthos of acidic, oligotrophic waters in low conductivities.



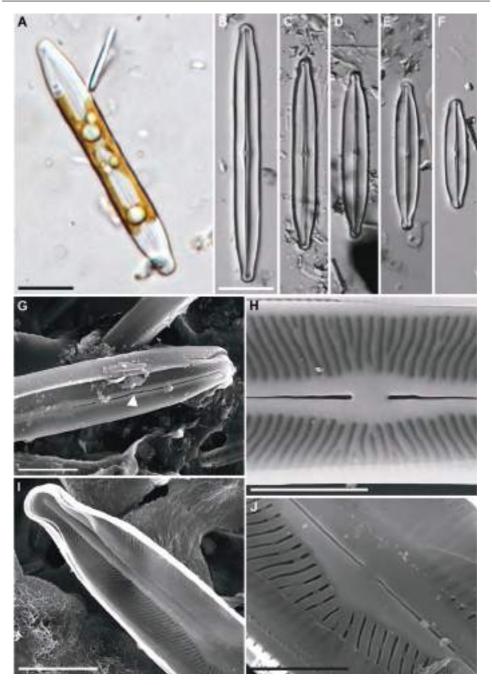


Fig. 142. *Kobayasiella* spp. **A-F.** LM. **A.** Living cell. **B-F.** Cleaned valves. **G-J.** SEM. **G-H.** External view of valve, note kink in the raphe (arrow). **I-J.** Internal view of valve.

Scale bars = 10 μ m (A-F), 5 μ m (G, I), 3 μ m (H), 2 μ m (J).

Mayamaea Lange-Bertalot 1997

Type species: Mayamaea atomus (Kützing) Lange-Bertalot

SYNONYM:

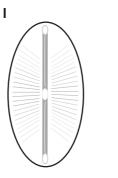
Navicula Bory 1822 pro parte

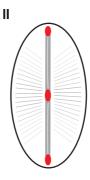
Characteristics – Cells biraphid, very small, elliptical with broadly rounded apices. Striae very difficult to discern under LM (Fig. 143: C-K) and also rather difficult to resolve with SEM. Raphe straight and simple (Fig. 143: C-K) carried in a sternum which, along with the striae mid-valve and near the valve margin, are usually the only structures which can be seen using LM. Slight swellings denoting the central and terminal nodules in the sternum at the central area and apices (II).

Plastid structure – Cells with one lobed plastid (Fig. 143: A-B), several lipid bodies scattered throughout the cell.

Identification of species – Species can be identified by cell size, cell shape, orientation and density of the striae as well as structure of the axial area.

Ecology – Cells solitary, free living and motile. Found in the benthos of alkaline eutrophic to hypereutrophic waters with moderate to high conductivities.





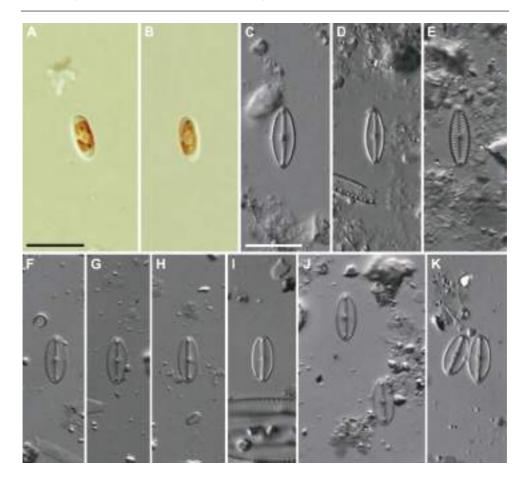


Fig. 143. Mayamaea spp. A-B. LM, living cells. C-K. LM, valve view of cleaned material. Scale bar = 10 μ m (A-K).

Navicula Bory 1822

Type species: Navicula tripunctata (O.F. Müller) Bory

Notes – Throughout this book many genera have *Navicula* listed as a synonym. For many years all diatoms cells exhibiting isobilateral symmetry and having a median raphe were placed into *Navicula* sensu lato. *Navicula* sensu stricto (in the strict sense) is now restricted to the former section lineolatae or those taxa having striae composed of linear areolae. Over the last 3 decades many taxa have been split off from *Navicula*; it is important to remember that this is an on-going process and that many more species currently in *Navicula* may in future be placed in other genera. In the interim, what may be termed as a 'catch all' genus has been established - *Naviculadicta* Lange-Bertalot 1994. This genus contains taxa without enough characteristics for description as a separate genus and which cannot be placed in *Navicula* sensu stricto. As more data (morphological or molecular) become available these taxa will be placed in new genera. We will not discuss or illustrate *Naviculadicta* in this volume as we do for the other genera as it not clearly a delimited entity.

Navicula Bory 1822

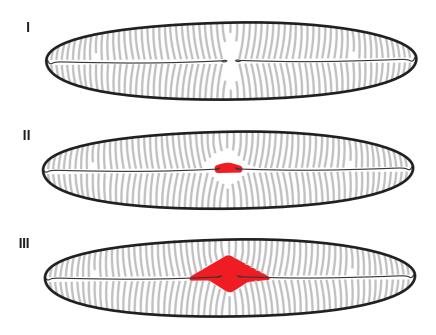
Type species: Navicula tripunctata (O.F. Müller) Bory

Characteristics – Cells biraphid, size, shape and apex structure variable. Striae discernable under LM (Fig. 145) and composed of a single row of linear areolae (lineolae; Fig. 146). In general striae are parallel mid-valve, become radiate and then often convergent towards the apices. Raphe carried in a sternum which in some taxa has a slight unilateral inflation (II) at the central nodule. The central area is variable in size and may not always be symmetrical (III).

Plastid structure – Cells with 2 plate-like chloroplasts, one along each side of the girdle (Fig. 144: B-E).

Identification of species – Species can be identified by cell size, cell shape, shape of the apices, structure and density of the striae as well as structure of the axial area and central area and the shape of the central and terminal raphe endings.

Ecology – Cells solitary, free living and motile. Found in the benthos of waters ranging from acidic to alkaline, oligotrophic to hypereutrophic and from low to high conductivities.



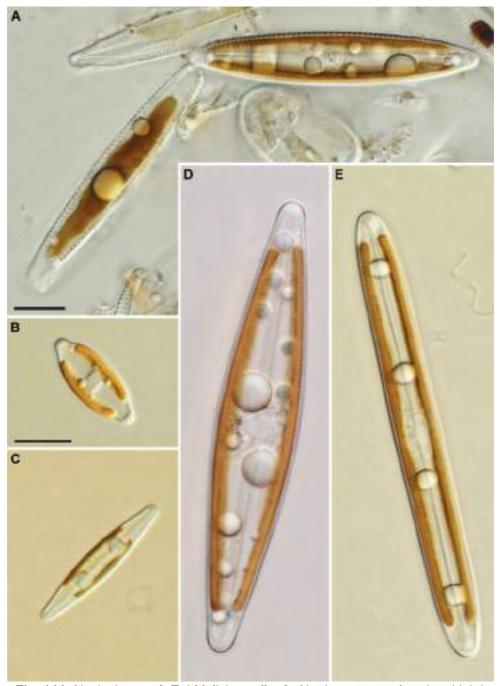


Fig. 144. *Navicula* spp. **A-E.** LM, living cells. **A.** *N. tripunctata*, valve view (right) and girdle view (left). **B-E.** Valve views. **B.** *N. radiosa* Kützing. **E.** *N. angusta* Grunow.

Scale bars = $10 \mu m (A-E)$.

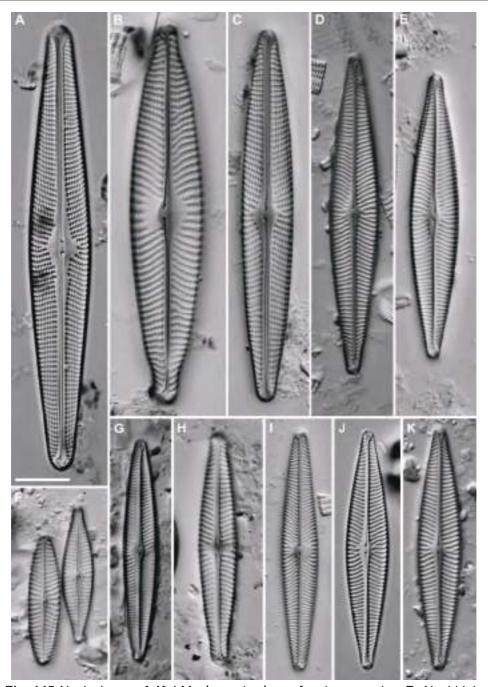


Fig. 145 Navicula spp. A-K. LM, cleaned valves of various species. B. N. viridula (Kützing) Ehrenberg. E. N. zanonii Hustedt. I. Navicula nielsfogedii J.C. Taylor & Cocquyt.

Scale bar = 10 µm (A-K).

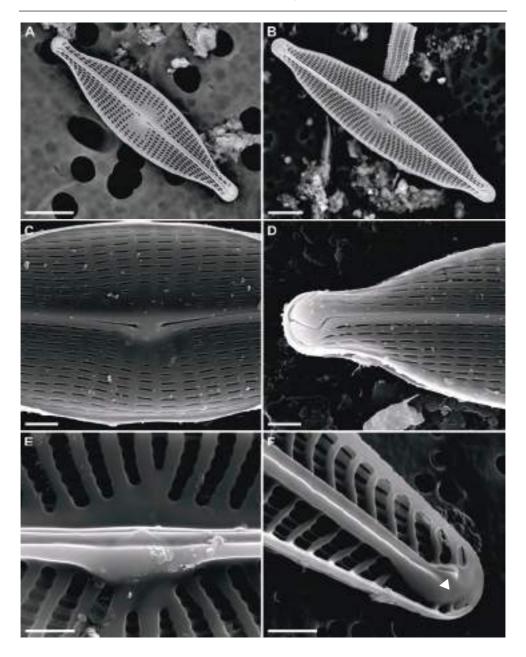


Fig. 146. Navicula spp. A-F. SEM. A-D. External view of valves. C. Detail of central raphe endings. D. Detail of terminal raphe ending. E-F. Internal view of valve. E-F. N. nielsfogedii, detail of central raphe endings (E) and terminal raphe ending (F), note helictoglossa (arrow). Scale bars = $5 \mu m$ (A-B), $1 \mu m$ (C-F).

Nupela Vyverman & Compère 1991

Type species: Nupela giluwensis Vyverman & Compère

SYNONYM:

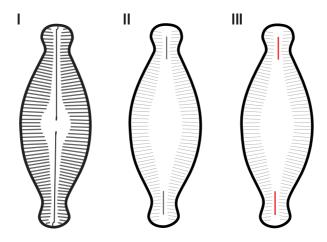
Navicula Bory 1822 pro parte

Characteristics – Cells isovalvar or heterovalvar, biraphid, small, elliptical to linear-elliptical, slightly asymmetric to the apical axis, with broadly rounded or protracted capitate or sub-capitate apices. Striae difficult to discern under LM (Fig. 147: E-R) composed of single rows of round or elongate areolae (Fig. 148: B-F). Raphe straight and simple (I; Fig. 148: C) extending on to the valve mantle, the opposite valve has short or very short and indistinct straight raphe branches which do not extend on to the valve mantle (III). Central area is asymmetrical and may be unilaterally expanded and may or may not reach the valve margins. Axial area often large (Fig. 148: B, D) and may be ornamented with valve face undulations.

Plastid structure – Cells with one plastid with lobes extending under the valve face (Fig. 147: A-D).

Identification of species – Species can be identified by cell size, cell shape, shape of the apices, orientation and density of the striae as well as shape of the central and axial areas.

Ecology – Cells solitary, free living and motile. Found in the benthos of slightly acidic to circumneutral waters with low conductivities.



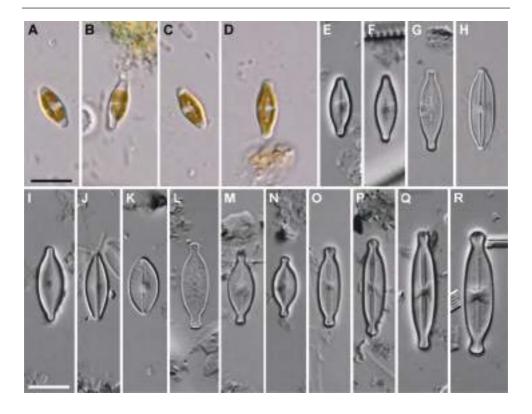


Fig. 147. Nupela spp. A-R. LM. A-D. Living cells. E-R. Valve views of cleaned material. Scale bars = $10 \mu m$ (A-R).

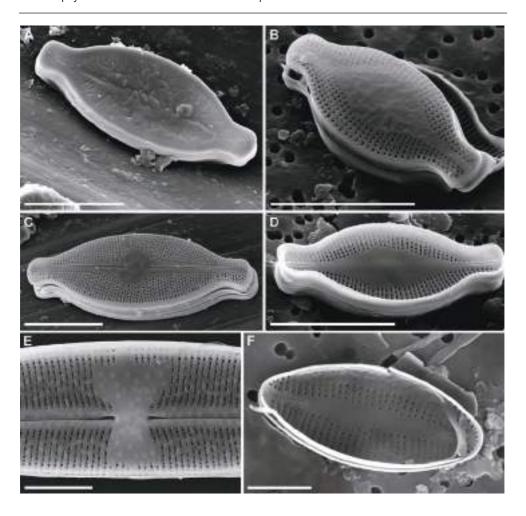


Fig. 148. *Nupela* spp. **A-F.** SEM. **A-C.** External view of valves, note short raphe branches (**B**). **D-F.** Internal view of valves, note short raphe branches (**D**, **F**). Scale bars = $5 \mu m$ (A-D), $2 \mu m$ (E-F).

Seminavis D.G. Mann 1990

Type species: Seminavis gracilenta (Grunow ex A.W.F. Schmidt) D.G. Mann

SYNONYM:

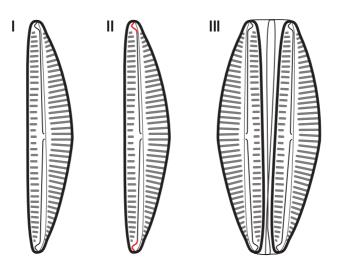
Amphora Ehrenberg ex Kützing 1844 pro parte

Characteristics — Cells **dorsiventral**, **biraphid**, straight ventral margin, curved dorsal margin with rounded apices. Striae discernable under LM (Fig. 149: A-C), composed of linear areolae only possible to resolve with SEM (Fig. 149: D-F). Raphe straight and simple (Fig. 149) carried in a sternum, terminal endings deflected to the dorsal side (II). Axial area and central area of different width and shape on dorsal and ventral sides. Differentiated from *Amphora* by the structure of the areolae and the plastids (naviculoid).

Plastid structure – Cells with 2 plate-like plastids, one along each side of the girdle.

Identification of species – Up till now only one species known from freshwaters of tropical Africa: Seminavis strigosa (Hustedt) Danielidis & Economou-Amili.

Ecology – Cells solitary, free living and motile. Found in the benthos of eutrophic waters with moderate to high conductivities.



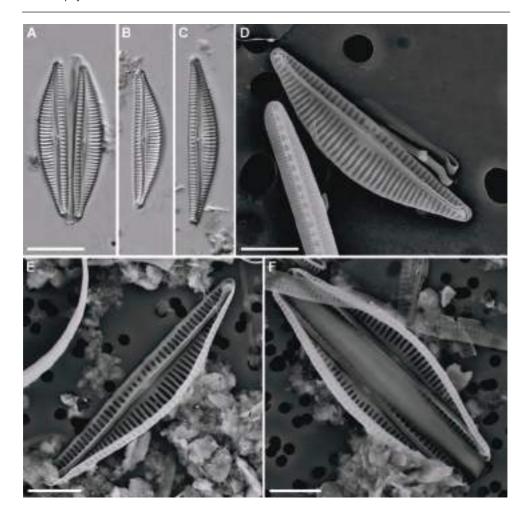


Fig. 149. Seminavis strigosa. A-C. LM, valve views. D-F. SEM. D. External view of valve. E-F. Internal view of valves. Scale bars = $10 \mu m$ (A-C), $5 \mu m$ (D-F).

Gyrosigma Hassall 1845

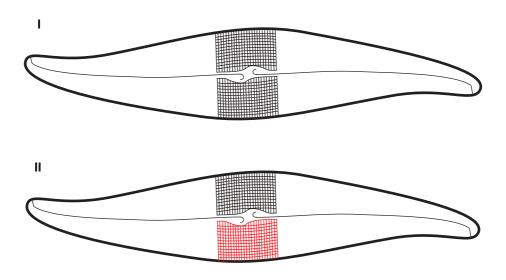
Type species: Gyrosigma hippocampus (Ehrenberg) Hassall

Characteristics – Cells **biraphid**, **sigmoid**, large to very large with rounded apices. Striae fine, transapical and longitudinal striae visible at right angles to each other (II; Fig. 151: C-D). Raphe sigmoid and simple (Fig. 150: C-D). Central area small and may contain special structures such as small silica ribs.

Plastid structure – Two plate-like chloroplasts sometimes with lobed margins lie along each side of the girdle (Fig. 150: A-B). Many lipid bodies scattered throughout the cell.

Identification of species – Species can be identified by cell size, cell shape, shape of the apices (degree of sigmoidality), structure and density of the transapical and longitudinal striae, structure of the central area as well as the shape and extent of the central raphe endings.

Ecology – Cells solitary, free living and motile. Found in the benthos of oligotrophic to eutrophic waters in both low and moderate conductivities.



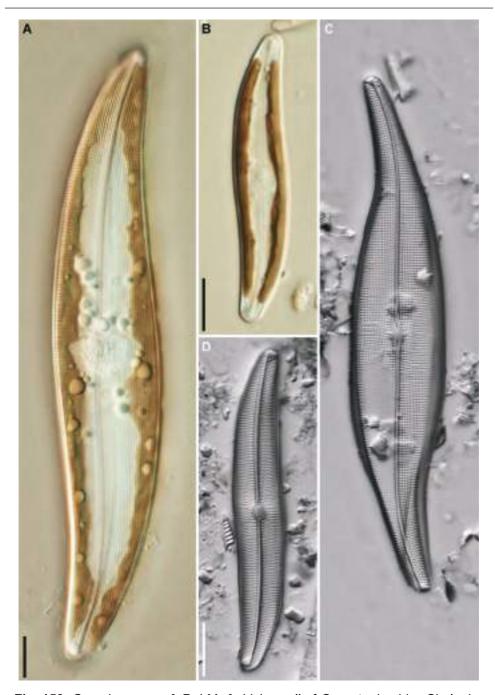


Fig. 150. *Gyrosigma* spp. **A-D.** LM. **A.** Living cell of *G. rautenbachiae* Cholnoky, note many lipd bodies. **B.** Living cell of *G. scalproides* (Rabenhorst) Cleve. **C.** Cleaned valve of *G. parkeri* (Harrison) Boyer. **D.** Cleaned valve of *G. scalproides*. Scale bars = 10 μ m (A-D).

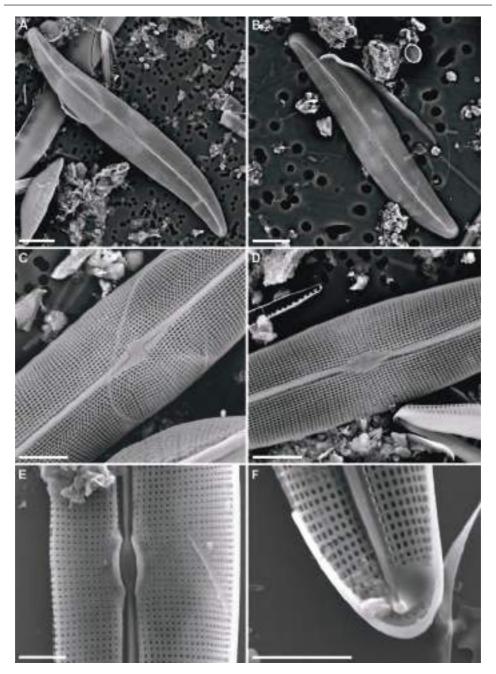


Fig. 151. Gyrosigma spp. A-F. SEM. A-D. External view of valves.

B. G. scalproides. C-D. Detail of central raphe endings. E-F. Internal view of valves. E. G. rautenbauchiae, detail of internal central raphe endings. F. Detail of internal terminal raphe ending and helictoglossa.

Scale bars = 10 μm (A-B), 5 μm (C-F).

Pleurosigma W. Smith 1852

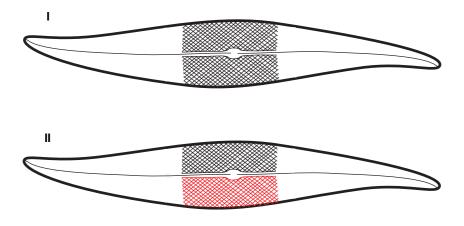
Type species: Pleurosigma angulatum (E.J. Quekett) W. Smith

Characteristics – Cells **biraphid**, **sigmoid**, large to very large with acutely rounded apices. Striae fine, transapical and longitudinal striae run diagonal to each other (II). Raphe sigmoid and simple (Fig. 152). Central area small, axial area very narrow.

Plastid structure – Two plate-like plastids, sometimes with lobed margins, lying along each side of the girdle (Fig. 152: A).

Identification of species – Species can be identified by cell size, cell shape, shape of the apices, structure and density of the striae as well as structure of the central area and the relative angle of the diagonal striae.

Ecology – Cells solitary, free living and motile. Found in the benthos of alkaline mesotrophic to eutrophic waters in moderate to high conductivities.



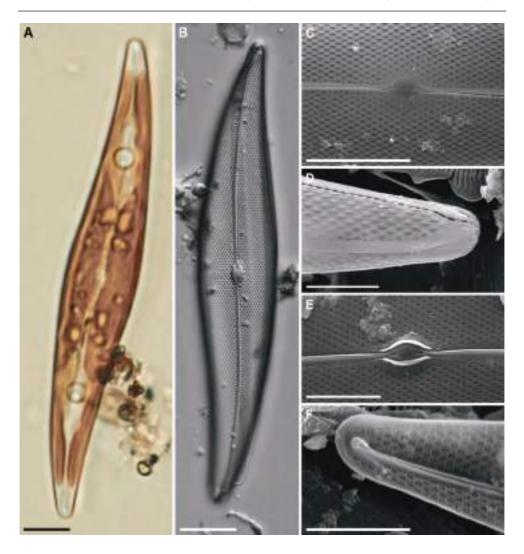


Fig. 152. Pleurosigma salinarum Grunow. A-B. LM. A. Living cell. B. Cleaned valve. C-F. SEM. C-D. External view of valve, detail of central raphe endings (C) and apex (D). E-F. Internal view of valve, detail of central raphe endings (E) and terminal raphe ending with helictoglossa (F). Scale bars = 10 μ m (A-B), 5 μ m (C-F).

Craticula Grunow 1868

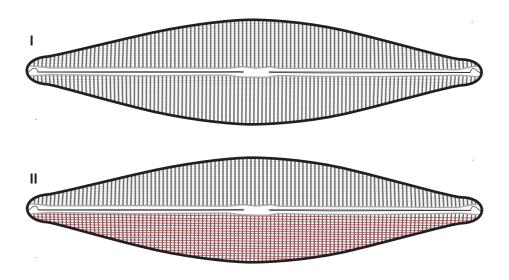
Type species: Craticula perrotettii Grunow

Characteristics – Cells **biraphid**, lanceolate with rostrate, capitate or broadly rounded apices. Striae parallel through the length of the valve. Areolae regularly arranged, very small and difficult to observe under LM (Fig. 154: A, B, D) but forming longitudinal striae (II). Cells of different species vary dramatically in size. Under certain conditions the cell forms a craticula (Fig. 1543: C), internal silica thickenings composed of a central rib and transverse ribs.

Plastid structure – Cells with one or two plastids on either side of the nucleus on each side of the girdle (clearly visible in large cells). Typically several small lipid droplets occur in the cytoplasm linking the plastids with one large droplet near to each pole (Fig. 153: C).

Identification of species – Species in this genus are distinguished based on cell size and shape as well as longitudinal and transverse striae density. The structure and shape of the central area can also be a useful characteristic.

Ecology – Cells solitary and motile. Found in the benthos of oligotrophic acidic water and extending into alkaline waters with high conductivity as well as very hard waters. Craticulae are formed when cells are exposed to high osmotic pressure.



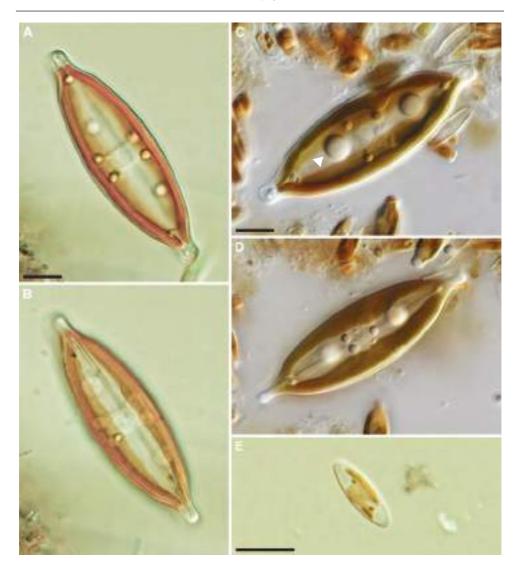


Fig. 153. Craticula spp. A-E. LM. A-B. Living cell of Craticula ambigua (Ehrenberg) D.G. Mann, valve view, different foci of same cell. C-D. Living cell of Craticula ambigua, valve view, different foci of same cell, note large lipid droplets (arrow). E. Craticula molestiformis (Hustedt) Mayama valve view.

Scale bars = 10 μm.

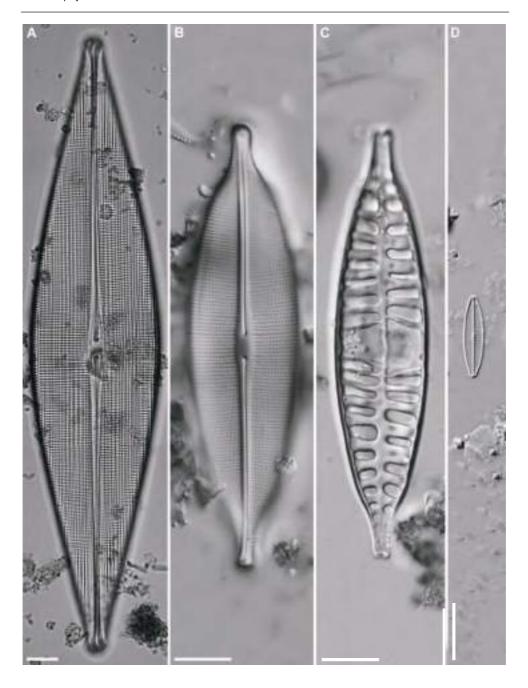


Fig. 154. Craticula spp. A-D. LM. A. Valve view of Craticula perrotettii. B. Valve view of C. ambigua. C. Craticula sp., a craticula. D. Valve view of Craticula submolesta (Hustedt) Lange-Bertalot.

Scale bars = 10 μm.

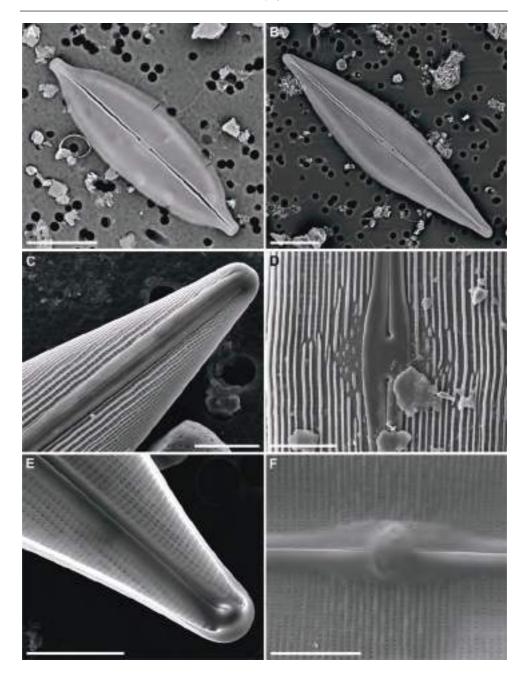


Fig. 155. Craticula spp. **A-F.** SEM. **A.** Valve view of Craticula ambigua. **B.** Valve view of C. cuspidata (Kützing) D.G. Mann . **C-F.** C. perrotettii, external view of terminal raphe ending (**C**), external view of central raphe endings (**D**), internal view of terminal raphe ending (**E**), internal view of central raphe endings (**F**). Scale bars = 20 μm (A-B), 10 μm (C-F).

Stauroneis Ehrenberg 1843

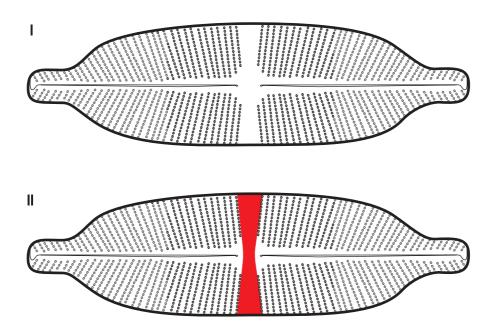
Type species: Stauroneis phoenicenteron (Nitzsch) Ehrenberg

Characteristics – Cells biraphid, cell may be large, elliptical to linear-elliptical and sub-capitate to capitate apices. Striae easily discernable under LM (Fig. 157) composed of a single row of round or elongate areolae (Fig. 158: A). Raphe carried in a sternum. Stauros present (II; Fig. 157; Fig. 158: D-E). Pseudosepta may be present at the apices (Fig. 158: C).

Plastid structure – 2 plate-like plastids extending under each valve (Fig. 156).

Identification of species – Species can be identified by cell size, cell shape, shape of the apices, structure and density of the striae, structure of the central raphe endings as well as structure of the central stauros and the presence/absence of pseudosepta.

Ecology – Cells solitary, free living and motile. Found mostly in the benthos of oligotrophic standing waters with low conductivities and also found in streams and sub-aerial habitats.



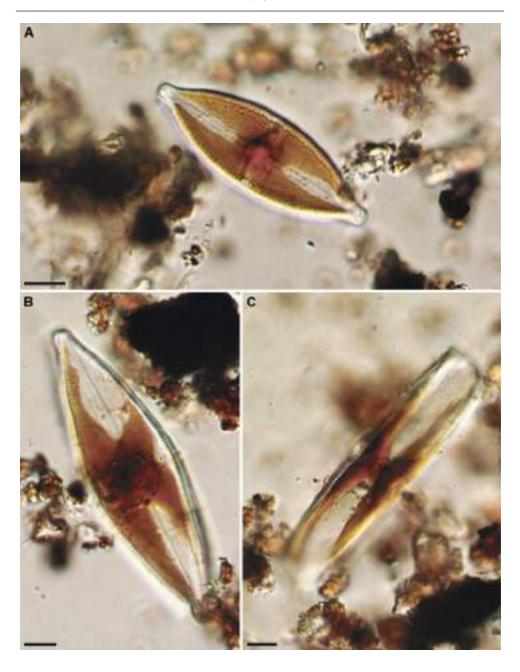


Fig. 156. Stauroneis spp. A-C. LM, living cells. A-B. Valve views. C. Girdle view. Scale bars = 10 μm .

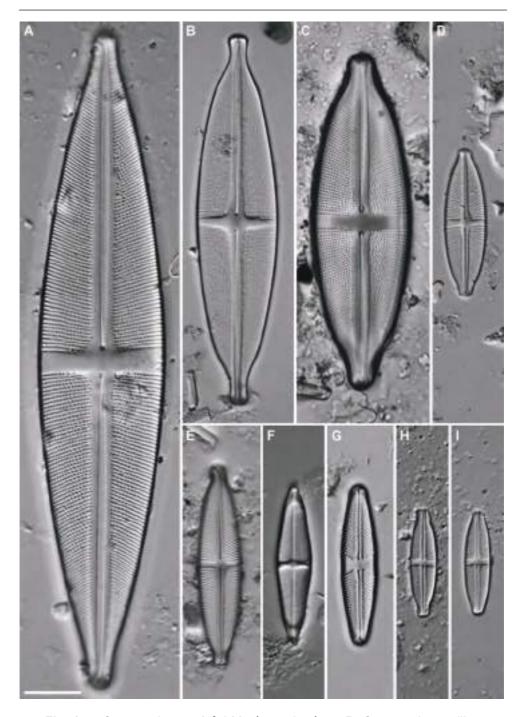


Fig. 157. Stauroneis spp. **A-I.** LM, cleaned valves. **B.** Stauroneis gracilior E. Reichardt. **H-I.** Stauroneis kriegeri R.M. Patrick. Scale bar = 10 μm (A-I).

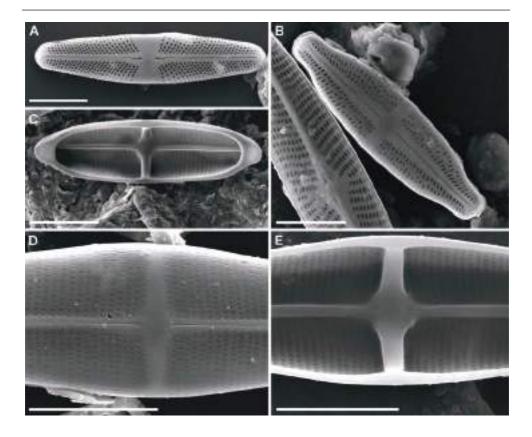


Fig. 158. Stauroneis spp. **A-E.** SEM. **A-B, D.** External view of valves. **B.** Stauroneis kriegeri. **C.** Internal view of valve, note the pseudosepta at both apices. **E.** Internal view of valve, detail of stauros. Scale bars = $5 \mu m$ (A-E).

Envekadea Van de Vijver, Gligora, F. Hinz, Kralj & Cocquyt 2009 Type species: *Envekadea hedinii* (Hustedt) Van de Vijver, Gligora, F. Hinz, Kralj & Cocquyt

SYNONYM:

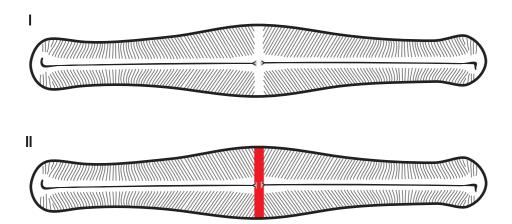
Navicula Bory 1822 pro parte
Stauroneis Ehrenberg 1843 pro parte

Characteristics – Cells **biraphid**, usually with expanded apices and expanded central region. Striae fine, strongly radiate in the mid-valve becoming strongly convergent near the apices. Raphe sigmoid, terminal raphe endings curved in opposite direction, golf club shaped under SEM, central raphe endings delta-shaped. Stauros may be present (II; Fig. 159).

Plastid structure - Cells with one H-shaped plastid.

Identification of species – Species can be identified by cell size, cell shape, shape of the apices, structure and density of the striae as well as structure of the central area.

Ecology – Cells solitary, free living and motile. Found in the benthos of oligotrophic to mesotrophic waters in both low and moderate conductivities.



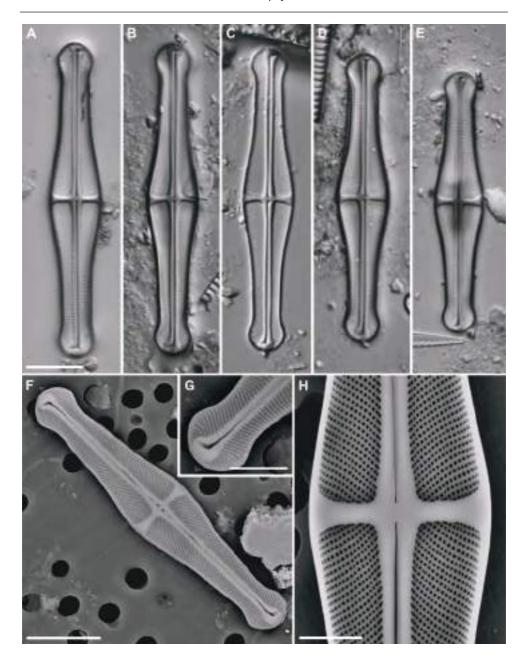


Fig. 159. Envekadea sp. A-E. LM, valve views. F-H. SEM, internal view of valve.
 G. Detail of apex, note golf club shaped terminal raphe ending. H. Detail of stauros, note delta-shaped central raphe endings.
 Scale bars = 10 μm (A-F), 5 μm (G), 3 μm (H).

Amphora Ehrenberg ex Kützing 1844

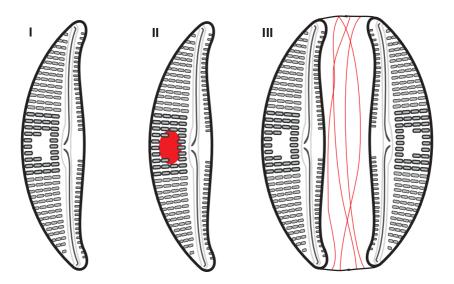
Type species: Amphora ovalis (Ehrenberg) Kützing

Characteristics – Cells biraphid, variable in terms of size and shape. Intact cells (i.e. both valves still joined by the girdle) are similar in shape to an orange segment, with the diatom valve faces being comparable to the faces of the orange segment, this is because cells have many more girdle bands on the dorsal side than on the ventral side (III; Fig. 161: A). The dorsal central striae are often separated by a thickened area of the valve known as a semi-stauros (II; Fig. 160: G-H; Fig. 161: D) absent in *Halamphora* Levkov. The striae on the ventral side of the valve are very short, composed of only a few areolae. In some species the areolae are clearly discernable under LM. Differentiated from *Halamphora* by the structure of the areolae (only visible under SEM).

Plastid structure – Single H-shaped plastid (Fig. 160: A). Lipid droplets (2-4) found towards the apex of each lobe of the plastid.

Identification of species – Species in this genus are distinguished based on cell size and shape and the shape of the apices. Striae density and angle relative to the **transapical axis** are also important characteristics to consider along with the size of individual areolae. The number of areolae on the ventral side of the valve is also important (IV) as well as the distance between the raphe and the ventral margin.

Ecology – Cells solitary, free living in the benthos of alkaline waters and occurring in a range of conductivities and trophic levels.



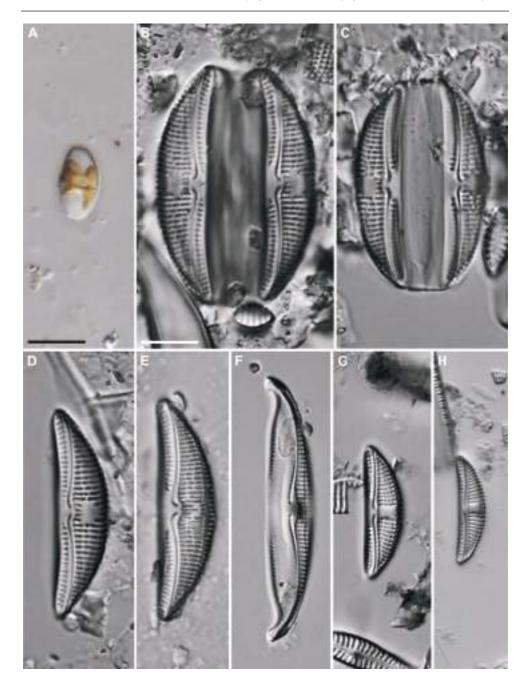


Fig. 160. Amphora spp. A-H. LM. A. Living cell. B-H. Cleaned valves. B-D. Amphora copulata (Kützing) Schoeman & R.E.M. Archibald. E. Amphora ovalis. Scale bars = 10 μ m (A-H).

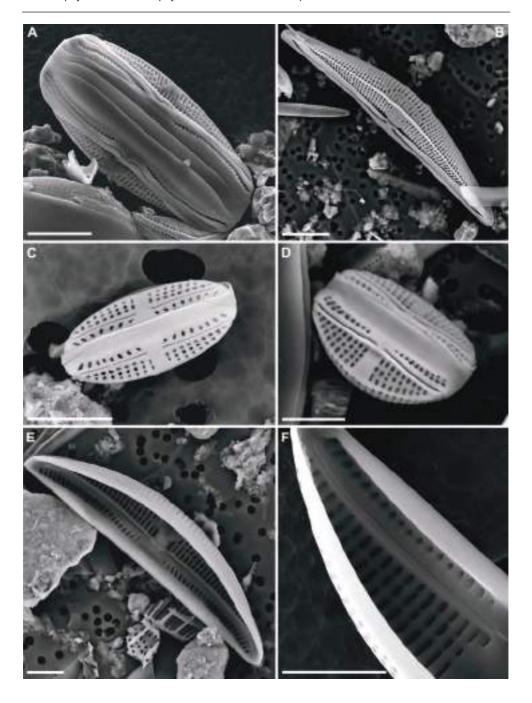


Fig. 161. Amphora spp. A-F. SEM. A-D. External view of valves. C-D. Amphora pediculus (Kützing) Grunow. E-F. Internal view of valve. Scale bar = 10 μ m (A-B), 5 μ m (C-F).

Halamphora (Cleve) Levkov 2009

Type species: Halamphora coffeaeformis (C. Agardh) Levkov

SYNONYM:

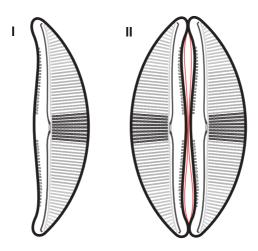
Amphora Ehrenberg ex Kützing 1844 pro parte

Characteristics – Cells **biraphid**, variable in terms of size and shape. Intact cells (i.e. both valves still joined by the girdle) are similar in shape to an orange segment, with the diatom valve faces being comparable to the faces of the orange segment, this is because cells have many more girdle bands on the dorsal side than on the ventral side (II; Fig. 163: A). Dorsal **semi-stauros** is absent. The striae on the ventral side of the valve are very short, composed of only a few areolae (Fig. 162: D, F-H; Fig. 163: C, E). In some species the areolae are clearly discernable under LM. Differentiated from *Amphora* by the structure of the areolae (only visible under SEM).

Plastid structure – Single H-shaped plastid (Fig. 162: A-B). Lipid droplets (2-4) found towards the apex of each lobe of the plastid (Fig. 162: A).

Identification of species – Species in this genus are distinguished based on cell size and shape and the shape of the apices. Striae density and angle relative to the **transapical axis** are also important characteristics to consider along with the size of individual areolae. The number of areolae on the ventral side of the valve is also important as well as the distance between the raphe and the ventral margin.

Ecology – Cells solitary, free living in the benthos. Occurs in a range of water quality with most species found at moderate conductivity and some species being specifically associated with high conductivity.



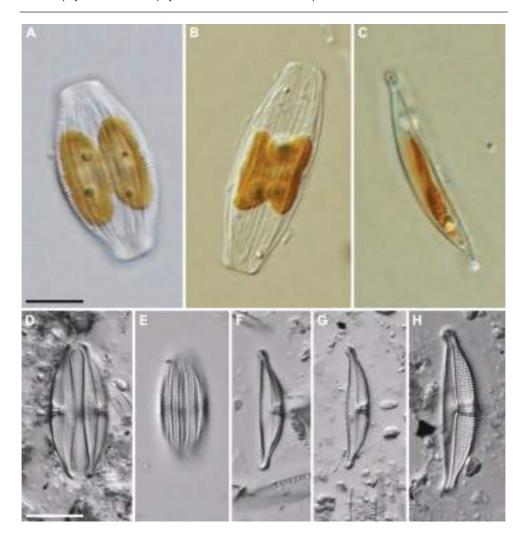


Fig. 162. Halamphora spp. A-H. LM. A-B. Living cells, girdle view. C. Living cell, valve view. D-H. Cleaned valves. D, F, G. Halamphora submontana Hustedt, valve view. E. H. submontana, girdle view.

Scale bars = 10 μm (A-H).